



# Orthohantaviruses Laboratory Detection

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# Diagnostic Challenges

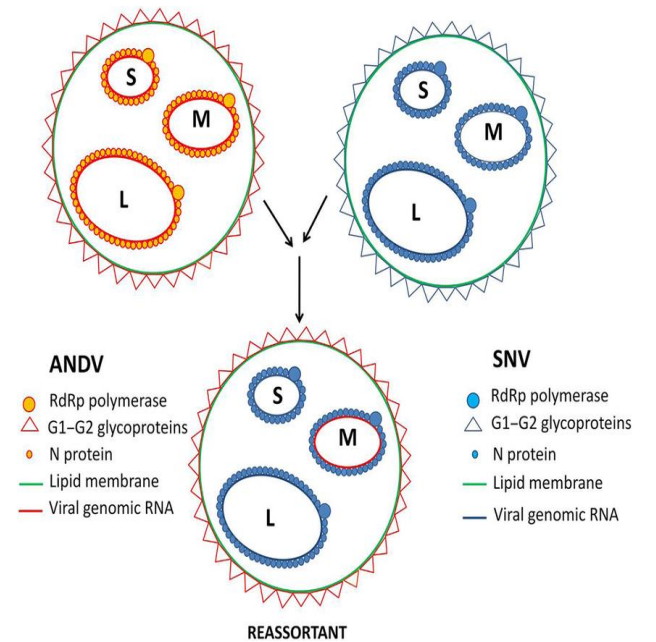
- Nonspecific early symptoms
- Mimics influenza/COVID/leptospirosis/dengue
- Rare disease in non-endemic settings

## Differential Diagnosis in Real-World Settings

Disease	Overlapping features	Differentiating tests
Leptospirosis	Renal failure, thrombocytopenia, fever	Leptospira PCR, blood culture, IgM
COVID-19	Interstitial pneumonia, fever, shock	SARS-CoV-2 RT-PCR
Sepsis of unknown origin	Shock, coagulopathy	Blood culture, procalcitonin, lactate
Rocky Mountain spotted fever	Fever, petechiae, thrombocytopenia	Rickettsia serology (Weil-Felix)
HCPS (ANDV)	Pulmonary oedema, cardiogenic shock, thrombocytopenia	Hantavirus IgM + RT-PCR

# Diagnostic Challenges

- Low viral load in acute phase: often <1000 copies/mL plasma
- Narrow timing windows: 3-7 days after symptom onset
- IgM antibodies appear late (day 5-14)
- Cross-reactive serology
- Genetic variability (reassortment, mutations) → false negatives in specific PCR
- Need for two independent methods (serology + molecular or sequencing)



# Laboratory testing of Andes virus (*Orthohantavirus andesense*) infection

Interim guidance

15 May 2026



- Preferred sample: **whole blood (EDTA)** – plasma has lower load
- serum, plasma, buffy coats can also be used Other specimens: urine, saliva/gingival swab in VTM, semen, CSF
- For fatal cases: **lung tissue** (fresh frozen or formalin-fixed) – PCR positive days after death

# Specimen Handling

## Preferred sample

## Storage

Whole blood  
Buffy coat  
Serum/plasma  
tissue

2–8°C short-term  
Delayed shipment –80°C/–20°C

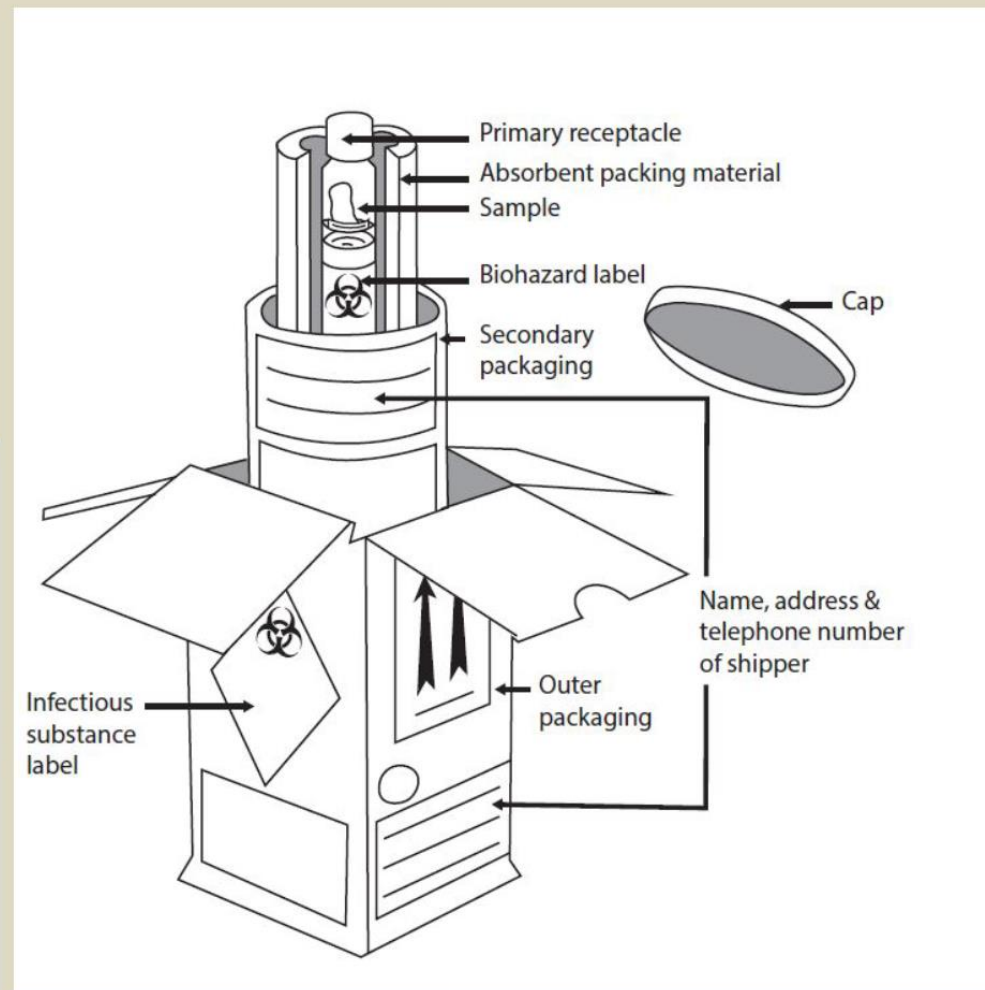
گیرنده:

تهران، میدان پاستور، خیابان ۱۲ فروردین،

انستیتو پاستور ایران، واحد تریاژ

هماهنگی پیش از ارسال نمونه:

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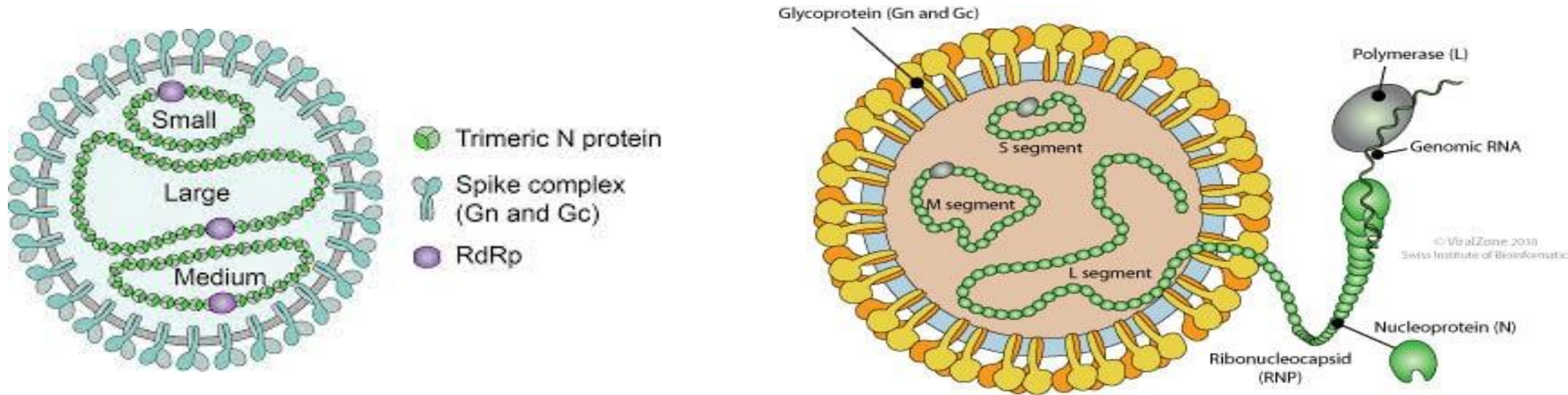


# Biosafety Considerations

- BSL2/BSL3 depending on risk assessment
- Risk-based approach
- Aerosol precautions
- BSC required before inactivation
- Inactivation protocols important
  - ANDV is Risk Group 3 (aerosol transmissible)
  - Before working in BSL-2, samples MUST be inactivated.

Chemical	Physical
- Methanol	- UV irradiation (254 nm, >30 min)
- Paraformaldehyde (4%)	- Heat inactivation (56°C for 30 min – may reduce PCR sensitivity)
- Acetone/Methanol (1:1)	
- Detergent-based lysis buffer (e.g., AVL buffer)	

# Genome Structure and Proteins



Vaheri A. Nature Reviews Microbiology. 2013

Segment	Protein	Key function	Role in diagnosis
S (~1.8 kb)	N	Nucleocapsid – highest expression, immunodominant	Main antigen in ELISA and primer target
S (ORF2)	NSs	Inhibits IRF3 phosphorylation, IFN- $\beta$ suppression	–
M (~3.6 kb)	Gn + Gc	Receptor binding ( $\alpha V\beta 3$ , $\alpha V\beta 1$ ), cell entry	Serology (less common)
L (~6.5 kb)	RdRp	Transcription/replication, endonuclease domain	Target for pan-hantavirus primers

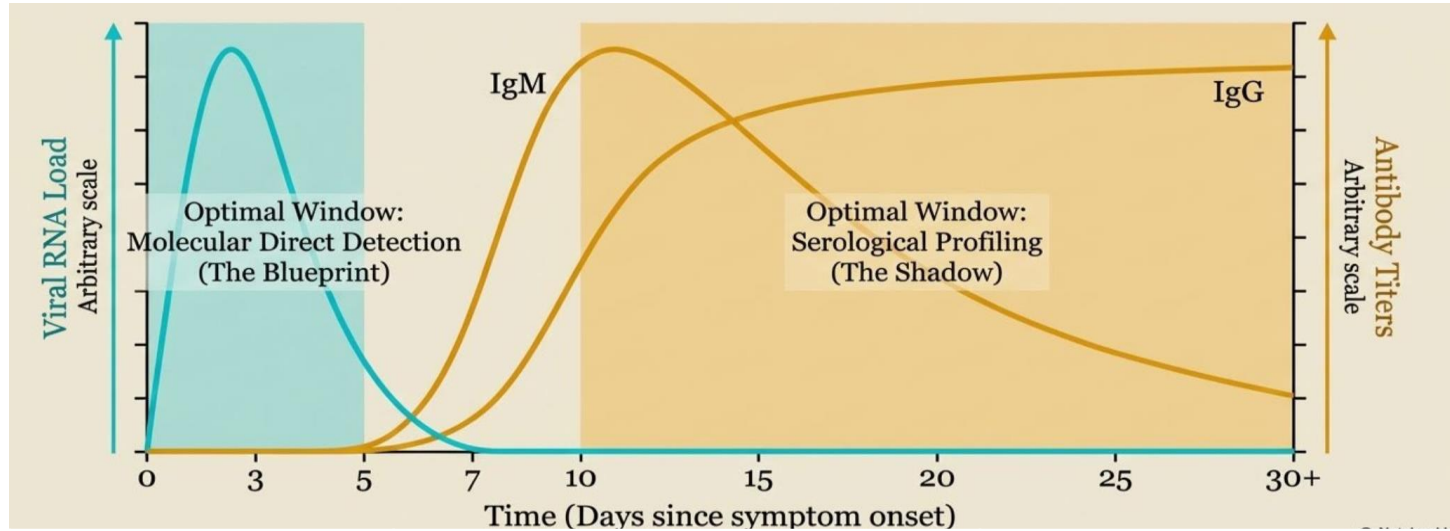
# Diagnostic Methods: Molecular (RT-PCR) and Serology (ELISA/IFA)

## Molecular (RT-PCR)

**Target:** Viral RNA (typically conserved S or L segments).

**Window:** Very early acute phase (prior to robust antibody response).

**Advantage:** Detects the virus before seroconversion; differentiates specific viral species and identifies early high viral loads linked to severe outcomes.



Stage	Detection Target	Method/Description
Early infection	Viral RNA	Specific primers or pan hanta approach high sensitivity needed
	IgM Antibody	Appears at symptom onset Acute infection marker
Peak protein expression	N protein	ELISA, immunoblot, IFA
Late convalescence	IgG antibodies	ELISA, FRNT / 4 fold rise

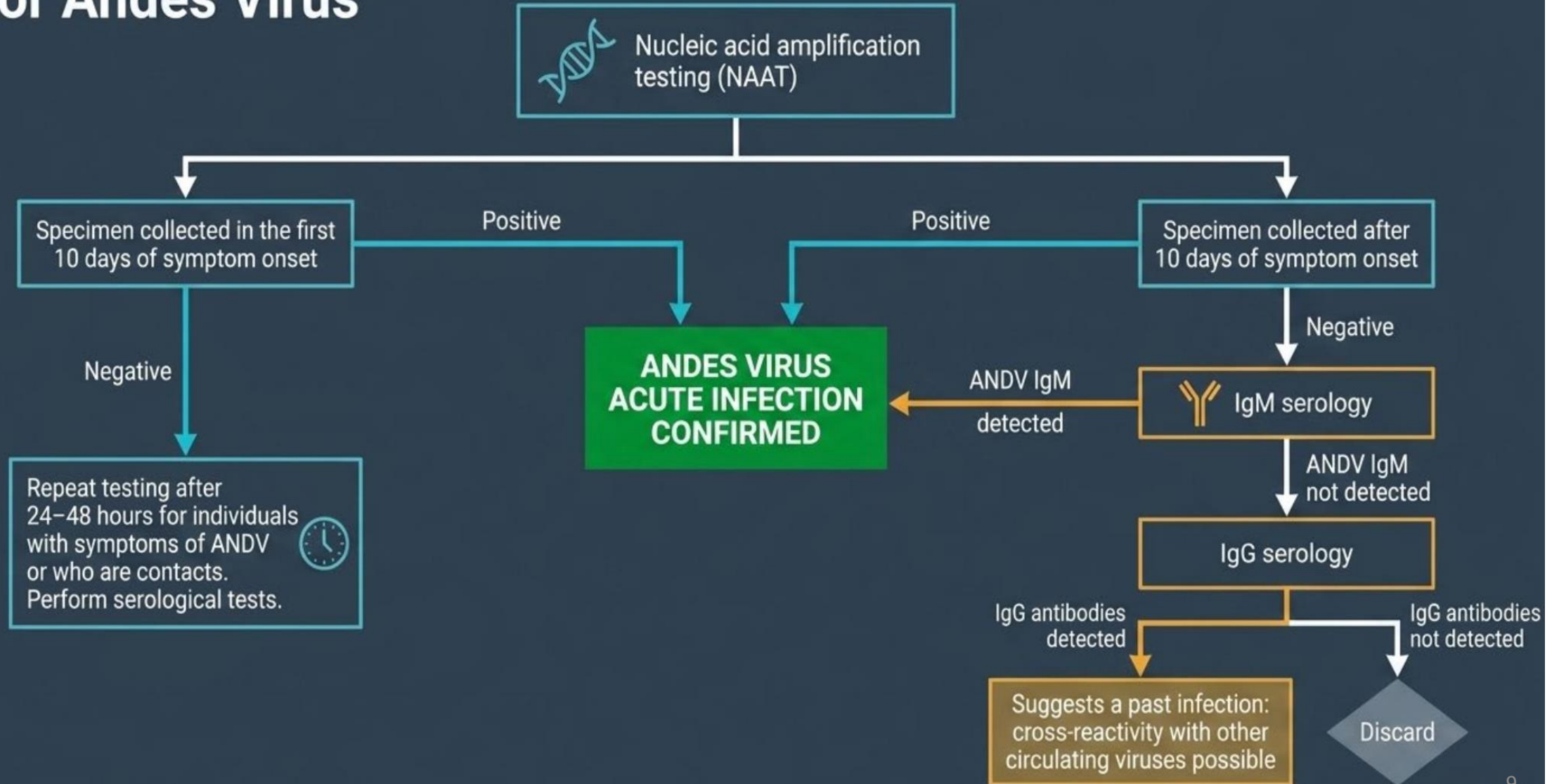
## Serology (ELISA/IFA)

**Target:** N-protein (highly conserved S-segment).

**Window:** Acute to Convalescent (IgM early, IgG long-term).

**Advantage:** The global gold standard. Immediate clinical confirmation upon symptom presentation.

# Comprehensive Diagnostic Algorithm for Andes Virus



# A single negative RT-PCR result is NOT sufficient to rule out ANDV infection.

Possible causes of false negative:

- Specimen collected too early (<3 days) or too late (>7-10days)
- **Low** viral load (<100 copies/mL plasma)
- Genetic mismatch (**reassortment, mutations** in primer binding sites)
- PCR inhibition or poor sample quality

What to do?

- If clinical suspicion remains high → **Repeat testing after 24–48 hours**
- Consider using a different target (e.g., **L segment vs. S segment**)
- Always run internal controls (positive, negative, inhibition)

# Commercial RT-PCR Kits

Kit	Manufacturer	Target	Viruses	Status
RealStar® Hantavirus-HFRS	Altona	RNA	DOBV, SEOV, HTNV, PUUV	RUO
RealStar® Hantavirus-HPS	Altona	RNA	ANDV, SNV	RUO
Sin Nombre Virus Real Time PCR	BioPerfectus	RNA	SNV	CE-IVD
Andes Virus Nucleic Acid Diagnostic Kit	Sansure	RNA	Andes	RUO

# Serological Kits: ELISA

Kit	Manufacturer	Antigen	Viruses	Status
Hantavirus (Hantaan) IgG/IgM ELISA	IBL	N protein (broad)	HTNV + cross-reactive	CE-IVD
Puumala IgG/IgM ELISA	Reagentia	N protein (PUUV-specific)	PUUV	CE
Hantavirus IgG/IgM ELISA	Euroimmun	N protein	Broad (Old & New World)	CE-IVD

# Essential for reliable hantavirus diagnostics.

## Common problems in European labs (2023 EQA):

- Detection of 100 copies/mL by PCR: only 68%
  - IgM serology agreement: 84%
  - Main issues: no internal control, poor melt curves, no replicates
- 
- Requirements:
    - Include positive, negative, and inhibition controls in every PCR run
    - Use RNase control for specimen integrity
    - Participate in EQA schemes (e.g., QCMD – Quality Control for Molecular Diagnostic s)
    - Follow ISO 15189:2022 (medical laboratories)

# Plaque Reduction Neutralization Test (PRNT)

- Principle: Measures antibodies that neutralize viral infectivity
- Sensitivity & Specificity: 100% (when performed correctly)
- Can **distinguish between closely related hantaviruses** (ANDV vs. SNV vs. HTNV)

## Limitations:

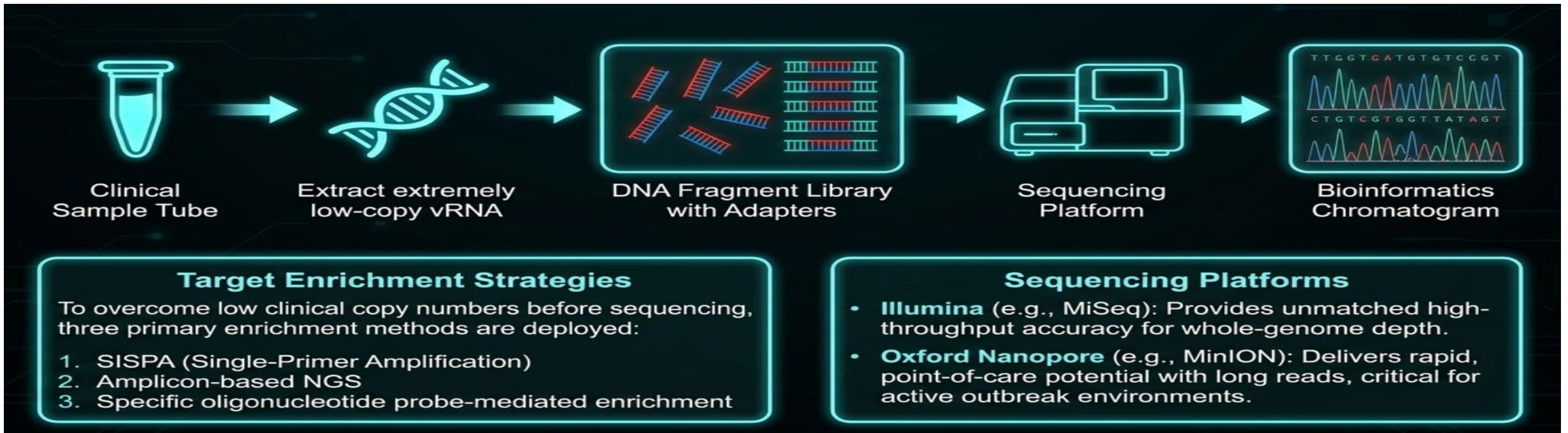
- Requires BSL-3 facility
- Time-consuming (5–7 days)
- Not for routine diagnosis – only reference laboratories

## Use case:

- Confirm equivocal ELISA/IFA results
- Discriminate cross-reactive antibodies in co-endemic regions
- Vaccine trials and research

# Next-Generation Sequencing (NGS) Methods

- Challenge: low viral load in human samples (often <100 copies/mL)
- Three main enrichment approaches (No et al. 2019; Park et al. 2021)



# DNA Microarray for Detection and Typing

- Third-generation **PathogenID v3.0** (Filippone et al. 2019)
- Tiled sequence covering entire S segment (assays 50 genotypes simultaneously)
- Advantage: can discriminate intra-species variants (e.g., ANDV Chile-9717869 vs. Arg-EP)
- Limitation: requires  $\geq 1000$  copies, 24 h turnaround
- Application: screening large rodent collections from hot spots

# This Will Not Be the Last Emerging Disease

- Next outbreak
- Delayed diagnosis leads to:
  - Missed window for early supportive care
  - Unrecognised transmission chains
- **Prevention:** Equipped laboratories, WHO guidance implementation, global collaboration, trained staff.



Thank you

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